

Kennesaw State University

DigitalCommons@Kennesaw State University

---

Master of Science in Integrative Biology Theses

Department of Ecology, Evolution, and  
Organismal Biology

---

Summer 8-4-2022

## Chemically Induced Phytoextraction of Lead (Pb) Contaminated Soil by Switchgrass (*Panicum virgatum* L.), assisted with BAP and NTA applications

Genna Hart

Follow this and additional works at: [https://digitalcommons.kennesaw.edu/integrbiol\\_etd](https://digitalcommons.kennesaw.edu/integrbiol_etd)



Part of the [Integrative Biology Commons](#)

---

### Recommended Citation

Hart, Genna, "Chemically Induced Phytoextraction of Lead (Pb) Contaminated Soil by Switchgrass (*Panicum virgatum* L.), assisted with BAP and NTA applications" (2022). *Master of Science in Integrative Biology Theses*. 87.

[https://digitalcommons.kennesaw.edu/integrbiol\\_etd/87](https://digitalcommons.kennesaw.edu/integrbiol_etd/87)

This Thesis is brought to you for free and open access by the Department of Ecology, Evolution, and Organismal Biology at DigitalCommons@Kennesaw State University. It has been accepted for inclusion in Master of Science in Integrative Biology Theses by an authorized administrator of DigitalCommons@Kennesaw State University. For more information, please contact [digitalcommons@kennesaw.edu](mailto:digitalcommons@kennesaw.edu).

# Chemically Induced Phytoextraction of Lead (Pb) Contaminated Soil by Switchgrass (*Panicum virgatum* L.), assisted with BAP and NTA applications

by  
Genna Hart

A Thesis Presented in Partial Fulfillment of Requirements for the Master of Science in Integrative  
Biology for the Department of Ecology, Evolution and Organismal Biology

Kennesaw State University  
1000 Chastain Road  
Kennesaw, GA  
August 2022

---

Thesis Chair: Dr. Sigurdur Greipsson  
Associate Professor of Biology  
Kennesaw State University

---

Committee Chair: Dr. Marina Koether  
Professor of Chemistry  
Kennesaw State University

---

Committee Chair: Dr. Thomas McElroy  
Associate Professor of Biology  
Kennesaw State University

## **Abstract**

The accumulation of lead (Pb) deposits in soil is a growing global concern. Phytoextraction of Pb-contaminated soil can be enhanced by soil chelation to increase the bioavailability of Pb. In the initial study nitrilotriacetic acid (NTA) 5mM, the alkyl polyglucoside (APG) Triton X-100 (2%), and nano-silica were applied to switchgrass (*Panicum virgatum* L.) growing in 5L pots of Pb-contaminated soil. The second experiment used 10mM NTA, the alkylpolyglycoside Triton X-100 (2%), foliar application of the plant growth regulator 6-benzylaminopurine (BAP) twice per week, Triple-12® nutrients, and Infuse™ a soil fungicide which were applied to switchgrass. Plants were grown in a controlled greenhouse environment in 5L pots with Pb-contaminated soil (5802.5 mg kg<sup>-1</sup>) collected from a former Superfund site in Cedartown, GA. Following harvest, plant material was dried, acid digested and chemically analyzed for lead (Pb), aluminum (Al) and iron (Fe) using an ICP-OES. Switchgrass treated with 5mM NTA showed an increased average Pb concentration of 858 mg kg<sup>-1</sup>, but this was further improved with applications of 10mM NTA, with an average concentration of 3196.5 mg kg<sup>-1</sup> in plants foliage. Combined applications of NTA and APG and foliar applications of BAP and nutrients resulted in the highest average Pb concentration of 5,355 mg kg<sup>-1</sup>. The results suggested that soil applications of 10 mM NTA and Triton X-100, Infuse™ and foliar applications of BAP could be a beneficial technique for phytoremediation of Pb contaminated soils.

## **Acknowledgements**

I would like to thank my supervisor and PI Dr. Sigurdur Greipsson for his knowledge, guidance, graduate program advice, and patience during my thesis work. Thank you for building the foundations for this project and encouraging me to learn additional skills along the way. I would also like to thank my committee chairs Dr. Marina Koether, for the use of the ICP-OES, her expansive knowledge and encouragement throughout this process and Dr. Thomas McElroy for their support. I would also like to thank Dr. McElroy for his assistance with my thesis proposal, his helpful advice about graduate school and for teaching foundational skills related to graduate coursework. Thank you, Dr. Melanie Griffin, Dr. Troy Mutchler, Dr. Dovid Kozlovsky, Dr. Susan Smith, Dr. Paula Jackson, Desiree Day, Dr. Joel McNeal, Dr. Mario Bretfeld, Dr. Daniel Ferreira and many others for your support and guidance.

Thank you, Will Beeson and my lab mate Kylie Stover, for your assistance in the lab. Many thanks to my undergraduate research assistants, Nailah Carter, and Nikki Shahi for support in lab and the greenhouse during this time. To my fellow graduate students, thank you for your encouragement and friendship, this cohort has made my time here at KSU even more enriching.

## Table of Contents

Abstract	1
Acknowledgements	2
List of abbreviations	5
<b>Introduction:</b>	
Sources of lead (Pb) soil contamination	6
Lead contamination and exposure	6
Risks of Pb in soil for humans	7
Switchgrass as a second-generation biofuel crop	8
Chelation and the phytoextraction process	8
Role of Arbuscular Mycorrhizal Fungi (AMF) in phytoremediation	9
AMF suppression	10
Utilizing the active growth regulator BAP	10
Chemical chelates used in phytoremediation	11
Foliar nutrient application	12
Focus of the study	12
<b>Materials and Methods:</b>	
Soil	13
Experiment 1	13
Experiment 2	14
Harvesting	15
Acid digestion and chemical analysis	15
Statistical Analysis	16
<b>Results:</b>	
Experiment 1: Effect of chemical chelates on switchgrass Pb, Al, Fe foliar concentrations	17
Experiment 2: Effect of APG and chemical chelates on switchgrass heavy metal concentration and shoot length	17
<b>Discussion:</b>	
Effects of chemical chelates on switchgrass growth	24

Chemical chelates and exploration of additional APGs	19
Effects of growth promoters and regulators	26
Dual applications of SNP and GA3	26
Salicylic acid and plant stress	27
Switchgrass as a biofuel and remediator	27
Previous studies on phytoextraction efficiency	28
<b>Future research directions</b>	29
<b>Conclusion</b>	31
<b>Statement of Integration</b>	31

## List of abbreviations

### Calculations:

ANOVA	Analysis of Variance
Fisher's LSD	Fisher's Least Significant Difference

### Chemical names:

APGs	Alkylpolyglucosides
BAP	6-Benzylaminopurine
EDTA	Ethylenediaminetetraacetic acid
GA <sub>3</sub>	Gibberellic acid
PGRs	Plant growth regulators
SA	Salicylic acid
SNP	Nitric oxide donor
Tergitol™ 15-S-9	Polyethylene glycol ether

### Elements:

Al	Aluminum
Pb	Lead
Fe	Iron
K	Potassium
N	Nitrogen
P	Phosphorus

### General:

AMF	Arbuscular Mycorrhizal Fungi
EPA	Environmental Protection Agency
ICP-OES	Inductively Coupled Plasma – Optical Emission Spectrometry

## **Introduction**

### **Sources of lead (Pb) soil contamination**

Accumulation of lead (Pb) deposits in soil is a growing concern due to the health implications and associated environmental damage [Anyanwu et al. 2018; Eiró et al. 2021]. Common causes of Pb contamination include run-off by factory operations, impurities in fertilizers used in farming, leaded gasoline emissions, smelting and mining operations [Zhuang et al. 2006].

Proximity to industries and former mining sites can also put communities at risk of Pb exposure [Mufalo et al. 2021]. Also, previous locations for smelting and recycling maintain high levels of Pb in nearby soil long afterwards [De Araújo and Do Nascimento 2010; Greipsson et al 2013; González-Chávez et al 2019]. Prior application of pesticides, including herbicides, previously aided in plant growth overall were identified as sources of heavy metal contamination [Alengebawy et al. 2021; Varshney et al. 2012]. Several studies have demonstrated that Pb contamination can come from a variety of sources and can affect the surrounding environment [Al-Sabbak et al. 2012; Chen et al. 2012; Fasinu and Orisakwe 2013; Wu et al. 2016].

### **Lead contamination and exposure**

Lead deposits in soil is a rising global concern in environmental analyses along with the dangerous future implications of soil contamination [Alsafran et al. 2021; Fasinu and Orisakwe 2013; He et al. 2015; Lwin et al. 2018]. Growing concentrations of heavy metal deposits in soils and their impact on the human body is a concern [Jan et al. 2015; Joosse 2022]. One drawback of commercial agriculture is the use of pesticides or herbicides which increases the number of heavy metals being deposited into soil [Alengebawy et al. 2021]. Also, sewage run off into residential areas in countries that are developing, and improper chemical dumping, can both have lasting effects on the soil [Brtnický et al. 2019]. As deposits continue to accumulate in the soil,

concerns have been raised regarding translocation of Pb into consumables and water sources [Alsafran et al. 2019; Chen et al. 2012; Fasinu and Orisakwe 2013; Zhuang et al. 2006]. Urban populations are most at risk for Pb exposure and the most effective and least invasive way to reduce this risk is to remove Pb from soils through phytoremediation [Filippelli and Laidlaw 2010]. The reduction of heavy metals in soil can benefit future crops and humans living in urban locations close to these contaminants [Filippelli et al. 2005; Zhuang et al. 2006].

### **Risks of Pb in soil for humans**

Recurring accumulating deposits of Pb in the soil or water systems has the potential to negatively impact its environment [He et al. 2015; Nazir et al. 2020]. Accumulation of Pb in soils has raised concerns about potential translocation of heavy metals from plant roots into the leaves [Zhuang et al. 2006]. Although federal organizations like the Environmental Protection Agency (EPA) have made guidelines to limit further Pb contamination and inform the public of the risks associated with Pb exposure, the current task at hand is to reduce the number and concentration of metals present in the soil [ACCLPP 2012; Filippelli and Laidlaw 2010]. In fact, there is no safe level of Pb exposure for humans, therefore any exposure to deposits of Pb in the environment has the potential to cause damaging human interactions later in life [Eiró et al. 2021, McFarland et al. 2022]. The ability to cross the blood-bone barrier is a major concern as Pb can remain present in the body for an unspecified length of time [Filippelli et al. 2005]. Urban populations have an increased risk of Pb exposure [Brtnický et al. 2019; Filippelli and Laidlaw 2010]. Long-term exposure to low-level Pb is associated with neurological damages which includes cognitive dysfunction in children [ACCLPP 2012; Canfield et al. 2006; Martinez-Finley et al. 2012; Paulson et al. 2019], along with Alzheimer's [Bakulski et al. 2012; Fathabadi et al.

2018] and Parkinson's diseases later in life [Chen et al. 2017]. Negative cognitive outcomes have been associated with children who had blood Pb levels of 10 mg/l or less [ACCLPP 2012; McFarland et al. 2022; Skerfving et al. 2015].

### **Switchgrass as a second-generation biofuel crop**

Switchgrass (*Panicum virgatum* L.) can grow in diverse environments and is a resilient plant once it is established [Fike et al. 2017]. The phytoremediation literature focuses mainly on grasses and similar monocots as they have a short growth process, are cost effective and are typically readily available in the region of interest [Chen et al. 2012; Johnson et al. 2015].

Switchgrass has several ideal characteristics coveted for phytoremediation: it has a high yield biomass and has shown resistance to heavy metal tolerance in previous studies [Chen et al. 2012; Gilly 2020; Greipsson 2011]. Switchgrass is utilized in the energy industry as a second-generation biofuel grass because it consistently yields acceptable harvestable biomass, has large ethanol conversion potential, and can grow in diverse ecosystems [Boyer et al. 2013; Hernández-Allica et al. 2008; Min et al. 2017]. Similar grasses easily show the effects of phytotoxicity or chlorosis rapidly and present a yellow or purple color as a positive indicator of heavy metal toxicity [Foy et al. 1978]. The outcome of the phytoextraction process has the potential to remove significant amounts of heavy metal contaminants from soil and restore the impacted soil for future uses [Greipsson et al. 2022; Huang et al. 1997].

### **Chelation and the phytoextraction process**

The removal of Pb-contaminants from soil can be accomplished through phytoextraction, an emerging cost-effective heavy metal remediation technique [Garbisu and Alkorta 2001;

Greipsson 2011]. Phytoremediation strategies include phytoextraction, using plants to remove heavy metals from contaminated soils [Greipsson 2011; Yan et al. 2020]. Phytoextraction usually requires acidification or chelation of soil which further facilitates the absorption and isolation of Pb in plant material [Foy et al. 1978]. Soil chelation may be improved based on the activation efficiency of the chelating agent used, but that is widely dependent on specific heavy metal interactions within soil [Liu et al. 2020]. Nitrilotriacetic acid (NTA), a derivative of EDTA (ethylenediaminetetraacetic acid) was the chelator used in this study. Successful chelation of Pb in soil has been shown to increase the bioavailability of Pb [Aderholt et al. 2017; Beavers 2016]. Increased motility of Pb in soil is often associated with higher concentrations of Pb in plants [Johnson et al. 2015]. One method of mobilization involves lowering the pH of the contaminated soil which allows Pb ions to mobilize and translocate ions into plant tissues, where it is stored in the leaves [Johnson et al. 2015]. Several other methods of Pb mobilization exist such as, oxidation or heating the soil of interest, but these processes can be expensive and may degrade the soil even further [Filgueiras et al. 2002].

### **Role of Arbuscular Mycorrhizal Fungi (AMF) in phytoremediation**

Symbiosis between the arbuscular mycorrhizal fungi (AMF) in the soil environment and the plant may be beneficial for long-term sustained growth and stress resistance [Cabral et al. 2015; Cruz-Paredes et al. 2021; Yang et al. 2016]. One benefit of AMF is the ability to aid in adaptive tolerance when plants undergo stress or exposure to a contaminated environment. [Azcón-Aguilar et al. 2002; Hovsepyan and Greipsson 2004]. A symbiotic site for chemical exchange and nutrient exchange is created between plants and AMF [van der Heijden et al. 2015]. The addition of plant hormones and uptake enhancers can be further regulated depending on the

specific AMF present [Greipsson 2011; Phielers et al. 2014]. Studies have suggested that the suppression of AMF can encourage the plant's heavy metal uptake from the soil and aid the plant in defense against metal toxicity [Hovsepyan et al. 2004; Perry et al 2012]. Other studies identified AMF tubules as a direct deterrent against Pb translocation from the roots into other foliage that could be isolated during harvest [Hildebrandt et al. 2007; Xu et al. 2014]. Soil fungicide may be used to arrest AMF activities to encourage phytoextraction of Pb into tissues of interest [Gilly 2020; Hart et al. 2022]. Further consideration of AMF use may be beneficial to future phytoremediation processes as alteration of the pH range may make plants more susceptible to heavy metal toxicity when AMF is suppressed [Coninx et al. 2017].

### **AMF suppression**

Infuse™ and its properties of pathogenic fungal resistance continue to be considered for future studies [Hovsepyan and Greipsson 2004]. Infuse™(benomyl) is a soil fungicide used to suppress AMF colonization within soil in phytoremediation research [Gilly 2020; Hart et al. 2022; Johnson et al. 2015]. Previous studies indicated that applications of another soil-fungicide benomyl prior to applying chemical chelates improved the uptake of Pb [Perry et al. 2012]. The application of Infuse™ reduced AMF colonization of root cells and treated plants showed an increase in Pb in switchgrass [Aderholt et al. 2017]. Additional studies specific to AMF suppression and its benefit to uptake of Pb and other heavy metals is suggested.

### **Utilizing the active growth regulator BAP**

Plant growth regulators are used to modify the plant's hormonal balance and stress endurance in reference to external mycorrhizal environments [Beavers et al. 2021; Evangelou et al. 2007].

Several studies have included the addition of growth regulators to further enhance the plant's ability to accumulate Pb [Aderholt et al. 2017; Gilly 2020; Perry et al. 2012]. A common application of exposing plants to growth regulators is foliar application or direct soil exposure [Benedetto et al. 2011; Matteo et al. 2015]. The growth regulator utilized in this experiment was Promalin®, which contained 6-benzylaminopurine (BAP). The BAP is a cytokine stimulator that has been shown to encourage plant bud growth and further stimulate multiple root growth via cell division [Bouré et al. 2019]. Combined applications of growth regulators can benefit the phytoremediation process and aid the plant's ability to uptake heavy metals [Aftab and Hakeem 2021].

### **Chemical chelates used in phytoremediation**

The addition of chemical chelates have been used to benefit the phytoremediation process and improve Pb availability. Varying concentrations of chelates were used depending on the soil, type of plants grown and focus of the phytoremediation study [Aderholt et al. 2017; Hovsepyan and Greipsson 2004]. Chelates are chemicals that can increase heavy metal availability during phytoremediation [Aderholt et al. 2017; Beavers 2016; Perry et al. 2012]. EDTA has been largely studied and identified as a successful chelator in several phytoremediation studies but has raised environmental concerns related to its limited biodegradability [Aderholt et al. 2017; Elliott and Brown 1986; Johnson et al. 2015; Gilly 2020; Greipsson 2011]. To limit potential environmental impacts, biodegradable chemicals other than EDTA including NTA and natural chelates like citric acid have been tested in phytoremediation studies as well [Chen et al. 2012; Shinta et al. 2021]. Citric acid has been identified as a natural chelate alternative to EDTA due to its ability to be broken down by microorganisms within soil [Aderholt et al. 2017; Beavers 2016;

De Araújo et al. 2010; Freitas et al. 2013]. A benefit to this limited amount of time spent in soil is a reduction in risk for groundwater contamination [Oviedo and Rodríguez 2003]. Future research regarding alternative chemical chelates and their associated applications may benefit the phytoremediation process [Aghelan et al. 2021; Chen et al. 2012].

### **Foliar nutrient application**

The foliar nutrient solution impacts plant growth by acting as a growth regulator to plants that undergo stress from heavy metal uptake [Sharma et al. 2020; Zottini et al. 2007]. Additional research focusing on salicylic acid and other alternative growth regulators in this field could benefit remediated plant's ability to uptake heavy metals during the phytoremediation processes [Beavers et al. 2021; Greipsson et al. 2021].

### **Focus of the study**

The focus of the experiments presented in this thesis was to determine the efficacy of Pb uptake through the phytoextraction process using switchgrass. The first experiment examined methods of Pb, Al and Fe uptake in switchgrass by evaluating the effect of chemical chelates (NTA) with the addition of permeating agent, Triton-X (100) and nano-silica. The second experiment determined the efficacy of Pb, Al and Fe uptake in switchgrass through the combined addition of chelating agents, phytohormones, surfactants, and a complete nutrient solution which included: nitrilotriacetic acid (NTA), Triple-12® (foliar nutrients), alkyl polyglycosides (APG ±Triton X-100) and the foliar application of Promalin® (includes BAP).

Objective 1 To examine methods of Pb uptake in switchgrass (*Panicum virgatum* L.) by evaluating the effect of chemical chelates 5mM Nitritoltriactic acid (NTA) with the addition of permeating agent, Triton X-100 or Nano silica.

Objective 2: To determine the efficacy of lead uptake in switchgrass (*Panicum virgatum* L.) through the addition of chelating agents, phytohormones, surfactants, and uptake agents including: 10mM Nitritoltriactic acid (NTA), Triple-12®, Triton X-100 (APG), Promalin® (includes BAP (6-benzyladenine)) and Infuse™ (fungicide).

## **Materials and Methods**

One hundred switchgrass seeds were allowed to germinate in a sandbox environment in the greenhouse for one month before the seedlings were transplanted into twelve, 5L pots containing Pb contaminated soil from a former Superfund Site in Cedartown, GA. Plants were grown in a controlled environment in Kennesaw State University's Science greenhouse. Plants were supplemented with white, fluorescent light for approximately 16 hours per day. Temperature (average 25 °C) and humidity was monitored and maintained at a near-constant level.

## **Soil**

Soil for this experiment was collected from a previous contaminated Superfund site in Cedartown, GA. Following the Agency for Toxic Substances and Disease registry (ASTR) reports in 2006, the soil remains Pb-contaminated with some high levels up to 260,000 mg kg<sup>-1</sup> present [ATSDR 2006]. Soil samples were randomly selected from the site and were

homogenously mixed prior to planting to ensure uniform heavy metal concentrations across experimental treatments.

## **Experiment 1**

In the first experiment, twelve switchgrass plants were grown with three plants in each treatment group. The goal of the first experiment was to determine the most effective alkyl polyglucoside (APG) with respect to Pb uptake.

Switchgrass seeds were germinated (as described above) and transplanted into 5L pots containing the Pb-contaminated soil from Cedartown, GA. In this study, chemically enhanced phytoextraction using the chelator nitrilotriacetic acid 5mM (NTA) with and without nano-silica or the alkyl polyglucoside (APG) Triton X-100 (TX 100) was tested [Hart et al. 2022]. At 56 dap the plants were subjected to four different experimental treatments: (1.) control (2.) nitrilotriacetic acid (NTA), (3.) NTA + nano-silica; and (4.) NTA + APG. The NTA (5 mmol kg<sup>-1</sup> soil) solution was made by mixing powdered NTA with an aqueous 10 N NaOH solution. The Triton X-100 (2%) was added to the NTA + APG solution in 100mL / 1L increments.

## **Experiment 2**

In the second experiment sixteen switchgrass plants were grown in the greenhouse. Switchgrass seeds were germinated (as described above) and transplanted into 5L pots containing the Pb-contaminated soil. Plants were grown under controlled conditions in the Kennesaw State University Research Greenhouse. Four replicated plants served as the control and did not receive

any phytohormone or APG treatments throughout the duration of the experiment. Four replicated pots were in each of the following treatments groups: (1.) BAP and NTA + Triton X-100 (2%), with foliar applications of Promalin® (1.67 mL/L), (2.) NTA 10mM and Triton X-100, (3.) Triple-12® (10mL/L) and NTA 10mM + Triton X-100, with foliar applications of Triple-12®, and (4.) control. The Triple-12® nutrient solution made by Growth Products Ltd. was used by mixing 5mL Triple-12® into 1L H<sub>2</sub>O. The Triple-12® foliar nutrient solution contained slow-release Nitrogen (N), and including 12% Nitrogen, 2% Phosphorous and 12% soluble Potash. The NTA (10 mmol kg<sup>-1</sup> soil) solution was made by mixing powdered NTA with an aqueous 10 M NaOH solution. The Triton X-100 (2%) was added to the NTA + APG solution in 100mL / 1L increments. The soil-fungicide Promalin® (propiconazole) solution, created by Valent BioSciences was made by mixing 1.67mL of aqueous Promalin® into 1L H<sub>2</sub>O. A complete nutrient solution containing: 332 mg L<sup>-1</sup> Nitrogen (N), 111 mg L<sup>-1</sup> Phosphorous (P) and 222 mg L<sup>-1</sup> K (Potassium) was supplied to all plants twice a week for 8 weeks until 64 DAP. All plants were watered with 200mL of water three times per week until 80 days after planting (dap). After 80 dap, plants with foliar applications were sprayed three times per week. The soil fungicide Infuse™ was applied after the 152 dap. The soil-fungicide was given to all plants for two weeks for suppression of symbiotic arbuscular mycorrhizal fungi. After 166 dap the 10mM NTA + Triton X-100 (2%) solution was applied to treatment groups 2, 3, and 4 three times per week until plants were harvested at 179 dap.

## **Harvesting**

Plants in all treatments and control groups were grown until fully mature. Plants were harvested following appearance of chlorosis and/or heavy metal toxicity and the presence of purple leaf

tips. The longest leaf, most affected leaves (visible chlorosis) and remaining leaves were collected from mature plants in all treatments. Harvested plant leaves were placed in paper bags, labeled, and dried in an oven at 65°C for 48 hours prior to acid digestion.

### **Acid digestion and chemical analysis**

Dried plant samples were weighed, placed in 100mL disposable acid digestion tubes (Environmental Express®, Inc) following a modified EPA method 3050 B and digested in a combined solution of 38% HCL (10.0mL) and 70% HNO<sub>3</sub> (10.0mL) [U.S. EPA 1996]. Plant samples were capped and placed in the solution for 24 hours until materials were mostly digested, and leaves appeared colorless. Digestion tubes were then placed in the Hotblock Express System™ and refluxed for 55 minutes at 90-95°C [U.S. EPA 1996]. Digestion tubes were allowed to cool to room temperature. Cooled digestion tubes had deionized water added to them up to the 100mL line and were vacuum filtered through 0.45-micron filters. Following filtration each sample was pipetted into 50 mL centrifuge tubes stored in a refrigerator until chemical analysis was performed by Dr. Marina Koether using inductively coupled plasma mass spectrometry (ICP-OES: Perkin Elmer Avio 200). Trace metals Al, Fe and Pb concentrations were identified in each sample collected and tested in triplicate against each standard. Standards for the metals of interest were generated by Dr. Marina Koether from Kennesaw State University's Department of Chemistry and Biochemistry.

### **Statistical Analysis**

Concentrations of Al, Pb and Fe were calculated and statistically analyzed through Analysis of Variance (ANOVA), average and standard deviation calculations, and Fisher's least significant difference (LSD) for post-hoc analyses. Excel's data analysis package along with the R studio's packages ggplot2, ggpubr, tidyverse, broom, AICcmoavg were used to perform the statistical tests of interest [Robinson et al. 2021; Rstudio Team 2020]. Lead data was transformed, and log transformation was employed via Excel. In terms of exploratory data analysis methods, normality tests were not applicable. The main statistical method used for analysis was one-way ANOVA through Excel. Data reorganization, determination of ANOVA type and data variance were verified in Rstudio with a combination of the following packages: ggplot2, ggpubr, tidyverse, broom and AICcmoavg [Rstudio Team 2020; Wickham 2020]. The ANOVA tables were calculated to identify interactions between variables. Fisher's (LSD) was used to determine the significance of p-values. Statistically significant p-values were accepted at  $p < 0.05$  and 5% (LSD) confidence levels,  $\alpha = 0.05$ .

## **Results**

### **Experiment 1: Effect of chemical chelates on switchgrass Pb, Al, Fe foliar concentrations**

Application of 5mM NTA + Triton X-100 resulted in statistically significant p-values of average Pb concentrations in the foliage when compared to the average Pb concentration from the control plants (Figure 1a). Plants in treatments receiving 5mM NTA showed an increase in average Pb, Al and Fe concentrations compared to plants in the control treatment (Figure 1a – 1c). Plants treated with NTA ± Triton X-100 had higher average Pb concentrations following statistical analysis compared to all other treatment groups (Figure 1a). Switchgrass treated with NTA + Triton X-100 had statistically significant p-values when compared to other treatments (Figure

4b). Overall, applications of 5mM NTA and Triton X-100 had a positive effect on Pb uptake. During the continued applications of NTA, the tips of plant leaves turned yellow and purple as the effect of metal induced chlorosis and toxicity were observed. None of the plants from any treatments had statistically significant p-values associated with Fe uptake (Figure 1c).

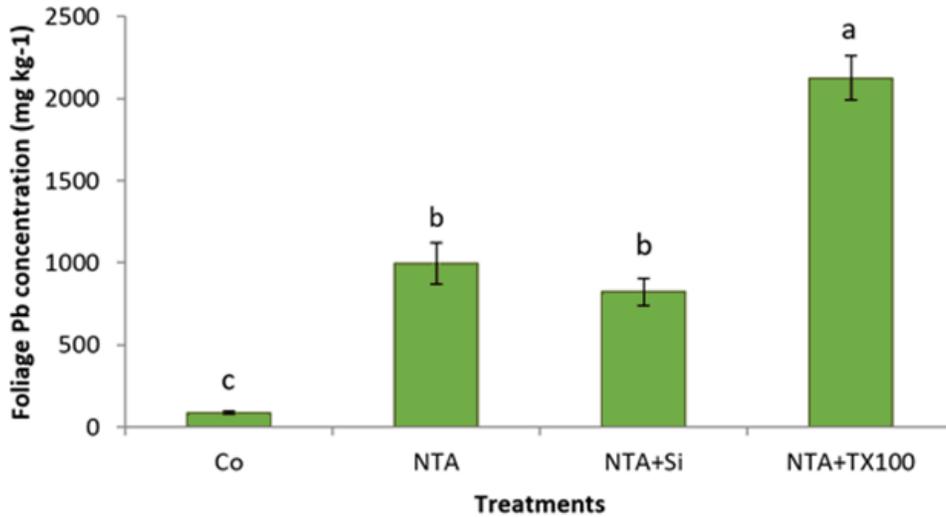


Figure 1a: Average Pb concentration (mg/L)  $\pm$ SD of *Panicum virgatum* as identified by ICP-OES analyses. Treatment group identifiers are as follows: Control = no treatment, NTA = 5mM nitriolotriacetic acid, NTA+Si = 5mM nitriolotriacetic acid + nano silica, NTA+TX100= 5mM nitriolotriacetic acid + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).

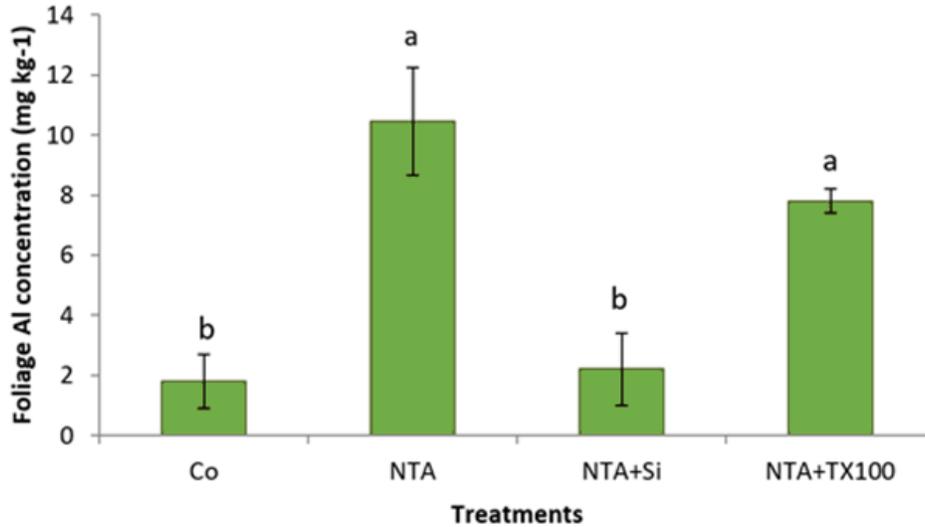


Figure 1b: Average Al concentration (mg/L)  $\pm$ SD of *Panicum virgatum* as identified by ICP-OES analyses. Treatment group identifiers are as follows: Control = no treatment, NTA = 5mM nitrilotriacetic acid, NTA+Si = 5mM nitrilotriacetic acid + nano silica, NTA+TX100= 5mM nitrilotriacetic acid + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).

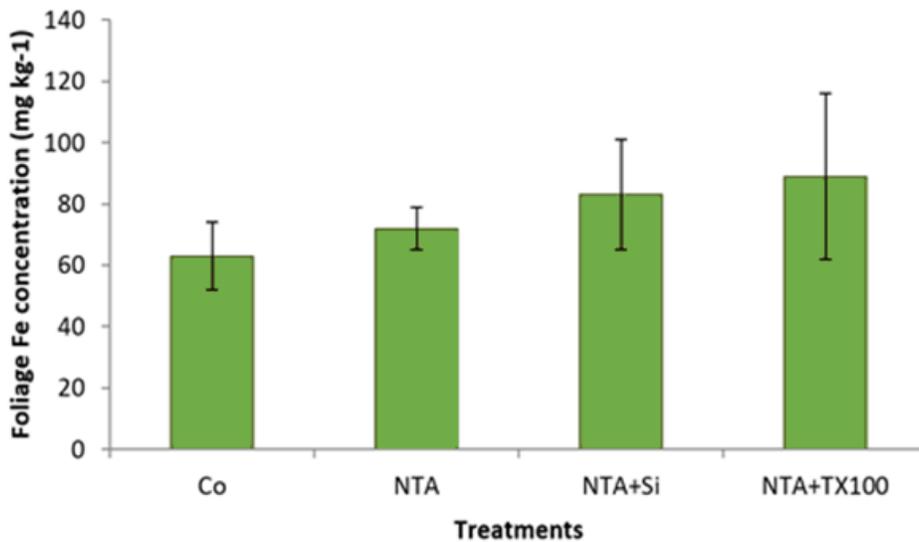


Figure 1c: Average Fe concentration (mg/L)  $\pm$ SD of *Panicum virgatum* as identified by ICP-OES analyses. Treatment group identifiers are as follows: Control = no treatment, NTA = 5mM nitrilotriacetic acid, NTA+Si = 5mM nitrilotriacetic acid + nano silica, NTA+TX100= 5mM nitrilotriacetic acid + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).



Figure 2: Untreated *Panicum virgatum* L. plants located on a wire benchtop in KSU's Research Greenhouse.



Figure 3: Isolated *Panicum virgatum* L. leaves displaying signs of chlorosis (yellowing).



Figure 4: Isolated *Panicum virgatum* L. leaves displaying early signs of necrosis.

## Experiment 2: Effect of APG and chemical chelates on switchgrass heavy metal concentration and shoot length

Two measurements were collected from each plant during harvest. Shoots were organized into two categories: longest leaf and leaves most affected by chlorosis. All plants treated with additional chemicals had longer average shoot lengths compared to the control plants (Figure 5). Plants with NTA + Triton X-100 had the longest average shoot length across all of the compared treatments (Figure 5).

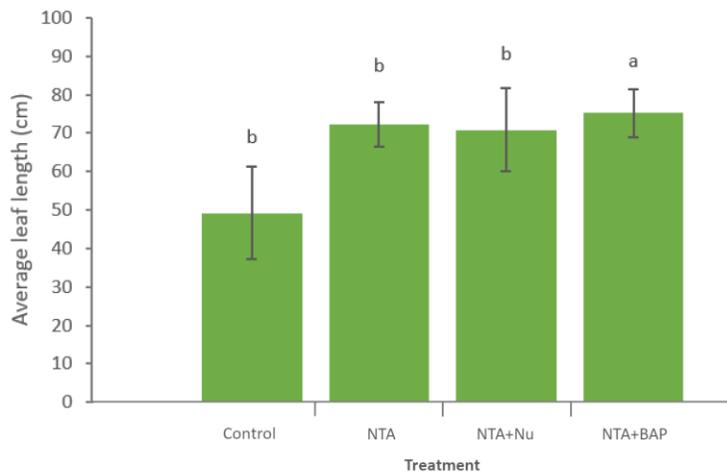


Figure 5: Average shoot length (cm) ( $\pm$ SD) of *Panicum virgatum* as measured by longest leaf length after 8 weeks of growth following applications of benzyl aminopurine (BAP), Triple-12® and 10mM nitrilotriacetic acid (NTA). Treatment group identifiers are as follows: NTA+Nu = Triple-12® + NTA 10mM + Triton -100, NTA = NTA 10mM + Triton X-100, NTA+BAP = Promalin® + NTA 10mM + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).

Treatment groups with 10mM NTA and BAP had the highest average Pb concentration across all treatments (Figures 6a-6c). Plants treated with 10mM NTA had a greater increase in average Pb concentrations than plants treated with 5mM NTA alone (Figure 6a, Figure 6a). One plant in the

10mM NTA and BAP treatment showed the highest Pb concentration of 10,820 mg kg<sup>-1</sup> Pb (Figure 6a). Experimental treatments of Triple-12®, BAP and NTA + Triton X-100 showed statistically significant p-values associated with Pb concentrations (Figure 6a). Lead analysis for plants treated with NTA + Triton X-100, BAP, and Triple-12® respectively, showed a p-value, p<0.01. Plants treated with NTA + Triton X-100, BAP and Triple-12® had a statistically significant p-value for Al, (p<0.01), when compared to the control plants. In reference to Fe, all treatments had a significant p-value, p<0.05. Combined applications of NTA, APG (Triton X-100) and BAP showed a significant increase in Pb, Al and Fe compared to the control (Figures 6a-6c). During the continued applications of NTA, the tips of plant leaves turned yellow and purple as the metal induced effect of chlorosis and metal toxicity were observed. Chlorosis was first observed following NTA applications after 170 dap.

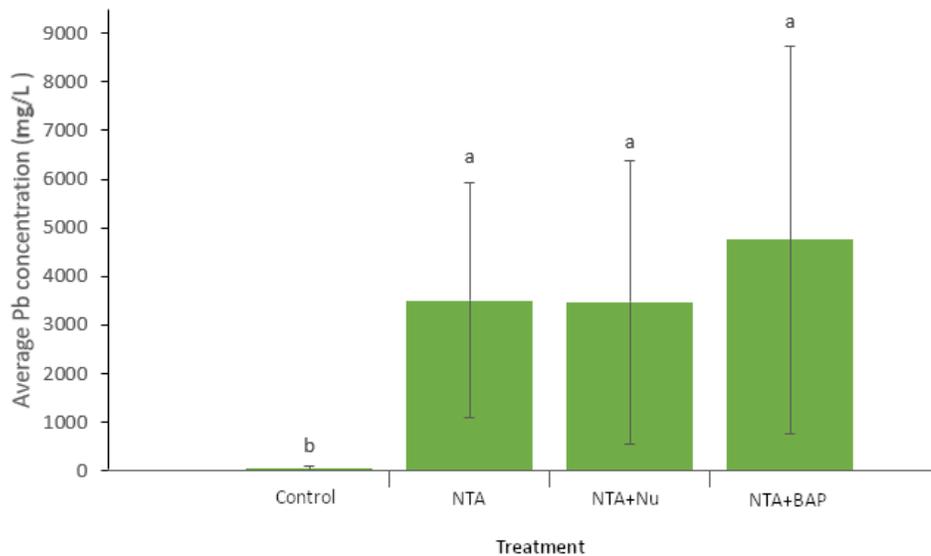


Figure 6a: Average Pb concentration (mg/L)  $\pm$ SD of *Panicum virgatum* as identified by ICP-OES analyses. Treatment group identifiers are as follows: NTA = NTA 10mM + Triton X-100, NTA+Nu = Triple-12® + NTA 10mM + Triton -100, NTA+BAP = Promalin® + NTA 10mM + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).

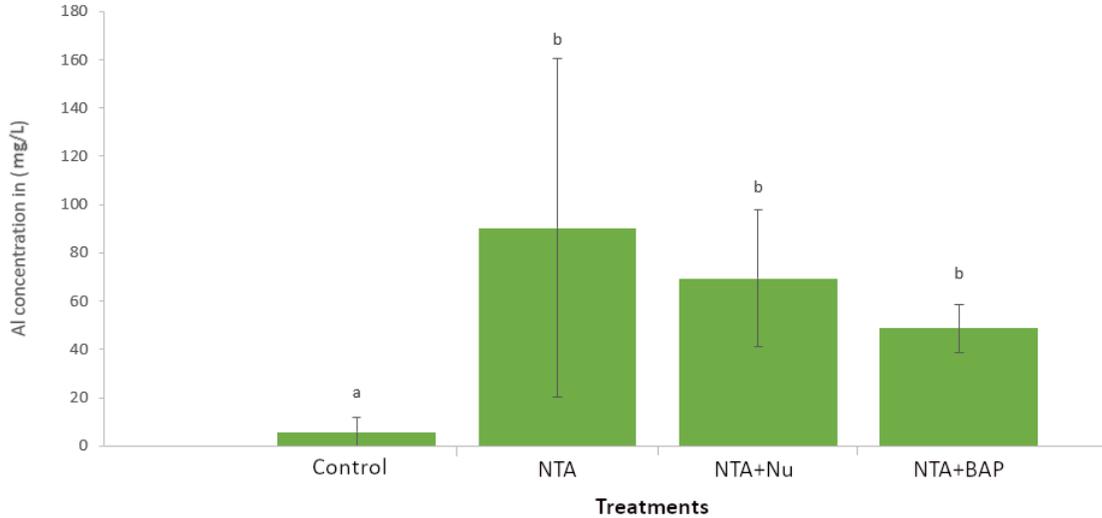


Figure 6b: Average Al concentration (mg/L)  $\pm$ SD of *Panicum virgatum* as identified by ICP-OES analyses. Treatment group identifiers are as follows: NTA = NTA 10mM + Triton X-100, NTA+Nu = Triple-12® + NTA 10mM + Triton -100, NTA+BAP = Promalin® + NTA 10mM + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).

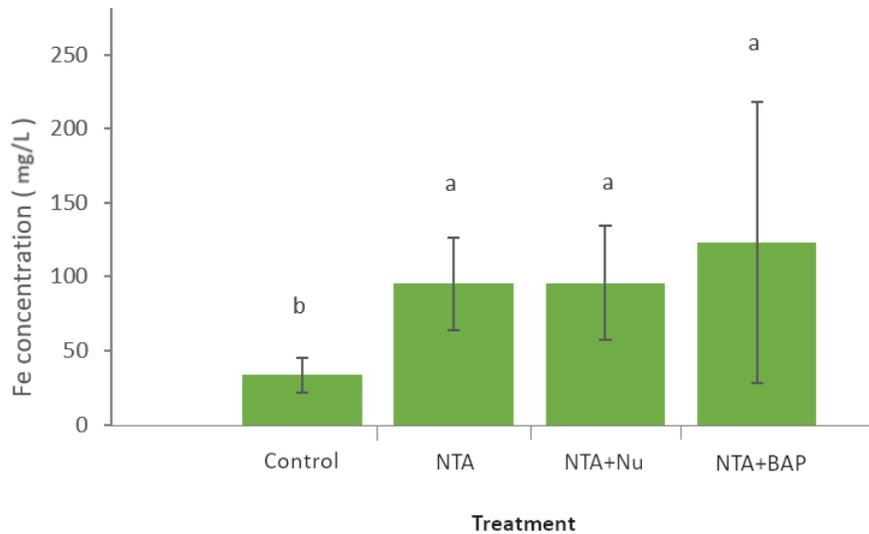


Figure 6c: Average Fe concentration (mg/L)  $\pm$ SD of *Panicum virgatum* as identified by ICP-OES analyses. Treatment group identifiers are as follows: NTA = NTA 10mM + Triton X-100, NTA+Nu = Triple-12® + NTA 10mM + Triton -100, NTA+BAP = Promalin® + NTA 10mM + Triton X-100. Means for columns with same letters are not significantly different ( $\alpha = 0.05$ ).

## **Discussion**

### **Effects of chemical chelates on switchgrass growth**

The addition of chemical chelates has been shown to enhance Pb uptake in switchgrass (*Panicum virgatum* L.) [Beavers et al. 2021; Evangelou et al. 2007; Hart et al. 2022]. The combined application of NTA +APG (Triton X-100), BAP® and Infuse™ may benefit future phytoextraction experiments. This was further supported by the largest Pb accumulation in one plant from 10mM NTA + APG and BAP treatment group (Figure 3a). Lead bioavailability increased with the application of NTA, and the use of this chelate could be beneficial as it degrades in remediated soil [Gilly 2020; Hart et al. 2022; Hu et al. 2017]. Further exploration of NTA and APG application can be investigated to determine their phytoremediation efficacy [Gilly 2020; Hart et al. 2022].

### **Chemical chelates and exploration of additional APGs**

Additional APGs can be explored in future studies as their applications may result in increased Pb uptake in switchgrass. Nitrilotriacetic acid (NTA) has been utilized to increase heavy metal bioavailability [Gilly 2020; Hart et al. 2022; Hu et al. 2017] The NTA utilization has shown to be effective in increasing the bioavailability of Pb when applied 10mM doses [Hart et al. 2022]. The NTA is more biodegradable in soil compared to its predecessor EDTA [De Araújo, J., and Do Nascimento, C 2010; Elliot and Brown 1989]. The environmental benefit, cost of NTA, and chelating effect are strong motivators to utilize this chemical [Hu et al. 2017]. Although EDTA has previously been utilized as an effective phytoextraction agent, its environmental persistence has encouraged researchers to utilize other chelates [Oviedo et al. 2003; Shinta et al. 2021]. Concerns with EDTA included Pb contamination of groundwater, resistance to breaking down in

soil, and EDTA remaining present in soil for a long time after chelation [Aderholt et al. 2017; Evangelou et al. 2007; Gilly 2020; Shinta et al. 2021]. The EDTA application was also associated with decreased plant growth and biomass [Meers et al. 2010]. Despite the current environmental concerns, EDTA has been widely utilized in previous studies as a successful chelating agent that increases the availability of Pb [Aderholt et al. 2017; Greipsson 2011; Johnson et al. 2015].

Several countries in the EU and Canada have limited EDTA use due to its harmful environmental impact and potential to impact aquatic organisms [EU Chemical Bureau 2004; Health Canada 2018]. Limitations of EDTA use including the impact of exposure and risks of extended environmental contamination was addressed in the EU's regulatory agencies [European Chemical Bureau 2004]. Continued assessment of this chemical and defined limits of its use to prevent environmental damage is an area for future research.

Further studies specific to the phytoextraction efficiency of combined applications of surfactants and chelating agents would be beneficial. Triton X-100 was the most effective alkyl polyglycoside (APG) identified in this study [Hart et al. 2022]. Dual applications of a surfactant and NTA improved the absorption, accumulation, and translocation of Pb when applied to triangular club-rush (*Scirpus triqueter* L.) [Hu et al. 2017]. The increased concentration from 5 mM NTA to 10mM NTA along with the APG Triton X-100 was associated with the largest Pb accumulation in switchgrass [Hart et al. 2022]. While applications of APGs and NTA could be successful in phytoremediation research, alternative ratios and timing of applications of these combined chemicals need to be examined further [Liu et al. 2020]. Future research might include

the results of APG applications when paired with the optimal concentration of NTA for different plant remediators and different soil types.

### **Effects of growth promoters and regulators**

Research on phytohormones, their effects on plant growth and Pb translocation has been outlined in several studies [Aderholt et al. 2017; Di Matteo et al. 2015]. Growth regulators like BAP have increased plant growth and assisted with heavy metal accumulation in plants in phytoremediation studies [Hart et al. 2022]. Previous studies have shown that exogenous applications of growth regulators were associated with an increase in Pb remediation [Gilly 2020; Greipsson et al. 2022]. When compared to other switchgrass related phytohormone treatments, BAP promoted the largest concentration of Pb in the foliage [Gilly 2020]. Although phytohormone use can benefit the phytoremediation processes, excessive use of these chemicals can be detrimental to plant health [Varshney et al. 2012].

### **Dual applications of SNP and GA<sub>3</sub>**

Previous study has shown that phytoextraction processes were improved with combined foliar applications of SNP (nitric oxide donor) and GA<sub>3</sub> (gibberellic acid) [Beavers et al. 2021].

Previous literature has addressed nitric oxide in further attempts to understand its role in cell signaling and the stress response [Khan et al. 2021; Namdjoyan et al. 2013; Neill et al. 2002].

The overall benefit of dual applications of SNP and GA<sub>3</sub> in the phytoremediation process is an area for further examination.

### **Salicylic acid and plant stress**

Salicylic acid (SA) has been utilized in studies as a plant hormone that assists with plants abiotic stress responses and has been shown to promote plant growth [Vicente et al. 2011; Zottini 2007]. Plants treated with SA displayed increased plant growth despite persistent heavy metal stress exposure [Horvath et al. 2007]. Zottini's study (2007) showed that some plants treated with SA were associated with reduced nitrogen reductase activity inhibitors. A decrease in nitrogen reductase inhibitors may result in increased switchgrass growth. Salicylic acid use was addressed in recent literature based on its ability to counteract heavy metal stress experienced by plants [Sharma et al. 2020]. Plants treated with low doses of SA tolerated more stress related to heavy metal uptake [Sharma et al. 2020]. Future studies utilizing SA applications and associated stress tolerance of plants undergoing phytoremediation is recommended.

A recent review article discussed salicylic acid (SA) use and its relationship with plant stress [Sharma et al. 2020]. The SA was used to help plants tolerate oxidative stress and endure the ongoing effects of heavy metal uptake [Zottini et al. 2007]. Exogenous applications of SA were beneficial for plants under stressful conditions [Sharma et al. 2020]. A recent study suggested that combined applications of SA alongside growth promotor DA-6 (Diethyl aminoethyl hexanoate) was associated with an increase in Pb uptake [Greipsson et al. 2022].

### **Switchgrass as a biofuel and remediator**

Switchgrass has previously been identified as a source of biofuel in the energy sector and successful remediator in phytoremediation literature [Balsamo et al. 2015; Min et al. 2017]. This is largely due to the plants amount of harvestable biomass, ethanol conversion potential and its ability to be grown in diverse climates as a remediator [Boyer et al. 2013; Hernández-Allica et al.

2008; Min et al. 2017]. Knowledge of plant biomass allowed researchers to determine a rate of remediation which would provide an environmental solution for Pb contaminated sites that are not currently remediated [Balsamo et al. 2015; Beavers 2016]. Continued research regarding switchgrass and its associated remediation biomass could solidify its use as an effective source of biofuel.

### **Previous studies on phytoextraction efficiency**

Previous experiments have tested the efficacy of phytohormones, varying chelating agents and Pb phytoremediation by switchgrass [Aderholt et al. 2017; Greipsson et al. 2022]. While Aderholt's (2017) use of EDTA did improve the bioavailability of Pb, applications with additional APGs showed greater Pb concentrations in the foliage of plants. The application of EDTA with the addition of phytohormone BAP and citric acid suggested more Pb uptake efficacy compared to EDTA alone [Aderholt et al. 2017;15, Johnson et al. 2015]. Phytohormone use and varying chelating agents paved the way for future experiments focused on switchgrass phytoremediation [Gilly 2020; Johnson et al. 2015]. Symbiotic use of AMF, several chelating agents including phytohormones, fertilizer treatments, and mycorrhizal treatments created a more tolerant environment for plant growth throughout the duration of the experiments [Gilly 2020; Hart et a. 2022]. Exogenous applications of plant growth regulators have been associated with an increase in phytoextraction efficiency [Beavers et al. 2021]. The switch to NTA from its predecessor EDTA was still associated with an increase in Pb bioavailability and a more effective soil degradation time following the experiment [Gilly 2020; Hart et al. 2022]. Alterations of the pH via sodium hydroxide suggests an effective manner to increase Pb uptake in phytoextraction processes [Foy et al. 1978]. The combined applications of phytohormones,

chelating agents, and AMF suppression treatments yielded higher levels of Pb in chemical analyses and leaves room for future studies regarding their effect on phytoremediation processes [Beavers et al. 2021; Hart et al. 2022] A more effective remediation process could help reclaim contaminated soils in a shorter time period and would limit heavy metal exposure to individuals in sensitive populations [Fasinu and Orisakwe 2013; Filippelli and Laidlaw 2010; Nissim and Labreccque 2021].

### **Future research directions**

Further study is required to identify the effects of increased concentrations of additional chemical chelates for switchgrass used in phytoextraction. Future experiments can specifically explore increased NTA applications to determine if the additional concentrations continue to be positively associated with Pb uptake. Successful removal of Pb from contaminated soils may benefit more sensitive human populations who are at risk of Pb exposure [Filippelli et al. 2005; Zhuang et al. 2006]. Switchgrass and similar monocots may be ideal models for phytoextraction efforts due to their short growth cycle, large biomass yield and resilience to phytotoxicity in the presence of Pb [Balsamo et al. 2015; Chen et al. 2012; Gilly 2020; Greipsson 2011]. Research encompassing combined applications of NTA, Triton X-100, foliar BAP, and Infuse™ yielding high Pb concentrations, and additional treatment combinations might improve upon past remediation efforts. Successful efforts of heavy metal uptake and Pb removal through phytoextraction could be applied through large scale practices to prevent further Pb accumulation and to remove current deposits from soil.

As demand for work in brownfield sites grows, so does its association with phytoremediation as these sites have high potentials for reclamation [Nissim and Labreccque 2021]. Brownfield sites utilize previously developed areas to continue development, or expansion [EPA 2020]. These redevelopments are further complicated if the soil at the site in question is contaminated with heavy metals [EPA 2018]. Specific government grant funding has been distributed towards remediation of eligible contaminated sites and there is a push to reclaim this soil [EPA 2018]. Often contaminated farm sites lack access to additional viable soil and remediation is a clear path to continued vegetation growth or utilization of sites [Yan et al. 2020]. There is a growing need for additional advances in effective phytoremediation practices to sustain the environment in affected cities [Anyanwu et al. 2018; Lee et al. 2021; Zhuang et al. 2006]. Continued phytoremediation research on the reclamation of brownfield sites and contaminated farmland may benefit the environment and associated redevelopment projects.

Several areas of continued research are recommended to strengthen the phytoremediation process and limit associated environmental concerns. The impact of additional APGs other than Triton X-100 could be researched in terms of phytoremediation including Tergitol™ 15-S-9 (Polyethylene glycol ether) [Dalton, 2019]. Varying concentrations of PGRs like benzyl aminopurine and stress tolerance from salicylic acid applications is another area for continued scientific research [Aderholt et al. 2017; Gilly 2020; Greipsson et al. 2022]. Utilization of paired SNP and gibberellic acid applications in previous studies had a positive association with plant growth and tolerance to chemical chelate applications and should continue to be researched [Beavers et al 2021]. Continued research focusing on the benefits of APGs, PGRs and chemical

chelate applications within phytoremediation could broaden areas of interest with current knowledge gaps.

## **Conclusion**

In conclusion, various approaches to phytoremediation may be beneficial to remediate contaminated soils. Different combinations of PGRs, APGs and chemical chelates may increase levels of heavy metal uptake. This study suggests that soil applications of 10 mM NTA and Triton X-100, Infuse™ and foliar applications of BAP could be a beneficial technique for phytoremediation of Pb contaminated soils. Lead uptake was amplified by the combined application of chemical chelates, growth regulators, and a soil fungicide. With the potential for Pb increase via phytoextraction, specific AMF and plant-soil interactions can be studied further. Phytoremediation studies involving switchgrass and arrested AMF activities can be studied further to determine the implications of suppression on phytoremediation. Increased Pb availability and the environmental impact of NTA, citric acid and EDTA use can be further assessed. Future research of additional chelating agents and chemical chelates may also be beneficial in the phytoremediation process of contaminated soils.

## **Statement of Integration**

This study integrated the following fields, biology, chemistry, sociology, and environmental sciences. Biology was used to assess plant growth regulators, surfactants, phytohormones and chelating agents to encourage Pb uptake in switchgrass plants. Concentrations of Pb, Al and Fe were isolated by chemical reflux reactions and later identified through chemical analysis and implementation of the ICP-OES. The chemistry of heavy metal interactions within soil and the

effect of chemical chelates were used to create the combined applications that experimental plants were treated with. Sociology was used in the study to identify how phytoremediation may benefit the environment and sensitive urban populations living near Pb-contaminated soils. Environmental science was implemented by identifying a need for phytoremediation and a reduction in heavy metal contamination in a specific soil sample.

## References

1. ACCLPP (Advisory committee on childhood lead poisoning prevention, of the centers for disease control and prevention). Low level lead exposure harms children: a renewed call for primary prevention; report to the CDCP; ACCLPP. 2012, 1-54.
2. Aghelan, N., Sobhanardakani, S., Cheraghi, M., Lorestani, B., and Merrikhpour, H. Evaluation of some chelating agents on phytoremediation efficiency of *Amaranthus caudatus* L. and *Tagetes patula* L. in soils contaminated with lead. *Journal of Environmental Health Science Engineering*. 2021, 19(1):503-514.  
<https://doi.org/10.1007/s40201-021-00623-y>
3. Al-Sabbak, M., Sadik, A., Savabi, O., Savabi, G., Dastgiri, S., and M. Savabieasfahani. Metal contamination and the epidemic of congenital birth defects in Iraqi cities. *Bulletin of Environmental Contamination and Toxicology*. 2012, (89):937-944.  
<https://doi.org/10.1007/s00128-012-0817-2>
4. ATSDR (Agency for toxic substances and disease registry). Public health assessment for Cedartown Industries, INC. Cedartown, Polk County, GA. U.S. department of health and human services. 2006; 1-48.
5. Al-Zurfi, S., and Al-Tabatabai, H. Aquatic plant (*Hydrilla verticillata*) roles in bioaccumulation of heavy metals. *Iraqi Journal of Agricultural Sciences*. 2020, 51(2):574-584. <https://doi.org/10.36103/ijas.v51i2.984>
6. Aderholt, M., Vogelien, D., Koether, M., and Greipsson, S. Phytoextraction of contaminated urban soils by *Panicum virgatum* L. enhanced with application of a plant growth regulator (BAP) and citric acid. *Chemosphere*. 2017,15-16:20-74.  
<https://doi.org/10.1016/j.chemosphere.2017.02.022>

7. Aftab, T., Hakeem, K.R. Plant Growth Regulators: Signaling under Stress Conditions, 1<sup>st</sup> ed.; Springer Nature Switzerland AG: Cham, Switzerland. 2021, 1-74:355-371.
8. Alengebawy, A., Abdelkhalek, S., Qureshi, S., and Wang, M. Heavy metals and pesticides toxicity in agricultural soil and plants: ecological risks and human health implications. *Toxics*. 2021;9(3):42. <https://doi.org/10.3390/toxics9030042>
9. Alsafran, M., Kamal, U., Hareb, A., and Muhammad, R. Ecological and health risks assessment of potentially toxic metals and metalloids contaminants: a case study of agricultural soils in Qatar. *Toxics*. 2021, (9), 2, 35. <https://doi.org/10.3390/toxics9020035>
10. Anyanwu, B., Ezejiolor, A., Igweze, Z., and Orisakwe, O. Heavy metal mixture exposure and effects in developing nations: an update. *Toxics*. 2018, (6)65:1-32. <https://doi.org/10.3390/toxics6040065>
11. Azcón-Aguilar, C., Jaizme-Vega, M.C., and Calvet, C. The contribution of arbuscular mycorrhizal fungi to the control of soil-borne plant pathogens. *Mycorrhizal Technology in Agriculture*. 2002:187-197. [https://doi.org/10.1007/978-3-0348-8117-3\\_15](https://doi.org/10.1007/978-3-0348-8117-3_15)
12. Bakulski, K., Rozek, L., Dolinoy, D., Paulson, H., and Hu, H. Alzheimer's disease and environmental exposure to lead: the epidemiologic evidence and potential role of epigenetics. *Current Alzheimer Research*. 2012, 9: 563–573. <https://doi.org/10.2174/156720512800617991>
13. Balsamo, R., Kelly, W., Satrio, J., Ruiz-Felix, M., Fetterman, M., Wynn, R., and Hagel, K. Utilization of grasses for potential biofuel production and phytoremediation of heavy metal contaminated soils. *International Journal of Phytoremediation*. 2015, 17(1-6):448-55. <https://doi.org/10.1080/15226514.2014.922918>

14. Beavers, A. Induced phytoextraction of lead from contaminated soils by *Panicum virgatum*, enhanced with EDTA, citric acid, benomyl, propiconazole and nitric oxide. 2016. Kennesaw State University, Kennesaw, GA, Summer 2016. Master of Science in Integrative Biology Theses.
15. Beavers, A., Koether, M., McElroy, T., and Greipsson, S. Effects of exogenous application of plant growth regulators (SNP and GA<sub>3</sub>) on phytoextraction by switchgrass (*Panicum virgatum* L.) grown in lead (Pb) contaminated soil. *Sustainability*. 2021, 13(19):10866. <https://doi.org/10.3390/su131910866>
16. Brandão, C., Valêncio, T., Bueno, M., Gonçalves, C., and Martinez, G. Accumulation and effects of copper on aquatic macrophytes *Potamogeton pectinatus* L.: potential application to environmental monitoring and phytoremediation. *Ecotoxicology and Environmental Safety*. 2018, (155): 117-124. <https://doi.org/10.1016/j.ecoenv.2018.01.062>
17. Bouré, N., Kumar, V., and Arnaud, N. The BAP module: A multisignal integrator orchestrating growth. *Trends in Plant Science*. 2019, (24)7: 602-610. <https://doi.org/10.1016/j.tplants.2019.04.002>
18. Boyer, C., Roberts, R., English, B., Tyler, D., Larson, J., and Mooney, D. Effects of soil type and landscape on yield and profit maximizing nitrogen rates for switchgrass production. *Biomass Bioenergy*. 2013, (48):33–42. <https://doi.org/10.1016/j.biombioe.2012.11.004>
19. Brtnický, M., Pecina, V., Hladký, J., Radziemska, M., Koudelková, Z., Klimánek, M., Richtera, L., Adamcová, D., Elbl, J., Galiová, MV., Baláková, L., Kynický, J., Smolíková, V., Houška, J., and Vaverková, M. Assessment of phytotoxicity, environmental and health

- risks of historical urban park soils. *Chemosphere*. 2019, (220): 678-686.  
<https://doi.org/10.1016/j.chemosphere.2018.12.188>
20. Cabral, L., Soares, C., Giachini, A., and Siqueira, J. Arbuscular mycorrhizal fungi in phytoremediation of contaminated areas by trace elements: mechanisms and major benefits of their applications. *World Journal of Microbiology and Biotechnology*. 2015, 31(11):1655-1664. <https://doi.org/10.1007/s11274-015-1918-y>
21. Calonne, M., Sahraoui, A. L. H., Campagnac, E., Debiane, D., Laruelle, F., Grandmougin-Ferjani, A., and Fontaine, J. Propiconazole inhibits the sterol 14 $\alpha$ -demethylase in *Glomus irregulare* like in phytopathogenic fungi. *Chemosphere*. 2012, 87(4):376-383
22. Canfield, R., Henderson, R., Cory-Slechta, D., Cox, C., Jusko, T., and Lanphear, B. Intellectual impairment in children with blood lead concentrations below 10 ug per deciliter. *New England Journal of Medicine*. 2003, 348: 1517–1526.  
<https://doi.org/10.1056/nejmoa022848>
23. Çelebi, Ş., Ekin, Z., and Eryiğit, T. Lead phytoremediation potential of hydroponically cultivated crop plants. *International Journal of Agriculture and Biology*. 2017, 19(5): 1141–1148. <http://doi.org/10.17957/ijab/15.0398>
24. Chaney, L., and Baklanov, I. Chapter five - Phytoremediation and phytomining. *Phytoremediation Advances in Botanical Research*. 2017, (83):189–221.  
<https://doi.org/10.1016/bs.abr.2016.12.006>
25. Chen, B., Lai, H., and Juang, K. Model evaluation of plant metal content and biomass yield for the phytoextraction of heavy metals by switchgrass. *Ecotoxicology and Environmental Safety*. 2012, (80): 393-400. <https://doi.org/10.1016/j.ecoenv.2012.04.011>

26. Chen, C., Chen, C. F., and Dong, C. Distribution and accumulation of mercury in sediments of Kaohsiung River mouth, Taiwan. *APCBEE Procedia*. 2012, 1:153-158.  
<https://doi.org/10.1016/j.apcbee.2012.03.025>.
27. Chen, H., Kwong, J., Copes, R., Tu, K., Villeneuve, P., van Donkelaar, A., Hystad, P., Martin, R., Murray, B., Jessiman, B., Wilton, A.S., Kopp, A., and Burnett, R. Living near major roads and the incidence of Dementia, Parkinson's disease, and multiple sclerosis: a population-based cohort study. *Lancet*. 2017, 389:718–726. [https://doi.org/10.1016/S0140-6736\(16\)32399-6](https://doi.org/10.1016/S0140-6736(16)32399-6)
28. Choudhary, V., Patel, M., and Pittman Jr., C., and Mohan, D. Batch and continuous fixed-bed lead removal using Himalayan pine needle biochar: *ACS Omega*. 2020, 5(27):16366–16378. <https://doi.org/10.1021/acsomega.0c00216>
29. Coninx, L., Martinova, V., and Rineau, F. Chapter four - mycorrhiza-assisted phytoremediation. *Advances in Botanical Research*. 2017, (83):127-188.  
<https://doi.org/10.1016/bs.abr.2016.12.005>
30. Cruz-Paredes, C., Diera, T., Davey, M., Rieckmann, M., Christensen, P., Dela Cruz, M., Laursen, K., Joner, E., Christensen, J., Nybroe, O., and Jakobsen, I. Disentangling the abiotic and biotic components of AMF suppressive soils. *Soil Biology and Biochemistry*. 2021 (159):1-11. <https://doi.org/10.1016/j.soilbio.2021.108305>
31. Cui, S., Zhang, T., Zhao, S., Li, P., Zhou, Q., Zhang, Q., and Han, Q. Evaluation of three ornamental plants for phytoremediation of Pb-contaminated soil. *International Journal of Phytoremediation*. 2013, 15(4):299-306. <https://doi.org/10.1080/15226514.2012.694502>

32. Dalton, M. An alternative to Triton X-100 for use in viral inactivation for recombinant factor viii. 2016. Atlantic Technological University. Galway, Ireland. Master of Science in Biopharmaceutical Science Thesis.
33. De Araújo, J., and Do Nascimento, C. Phytoextraction of lead from soil from a battery recycling site: the use of citric acid and NTA. *Water, Air, & Soil Pollution*. 2010, 211: 113–120. <https://doi.org/10.1007/s11270-009-0285-4>
34. Di Benedetto, A., and Pagani, A. Chapter 1 - Difficulties and possibilities of alternative substrates for ornamental bedding plants: an ecophysiological approach. Peat: formation, uses and biological effects. Nova Science Publishers Inc, UK. 2012;1-34.
35. Di Matteo, J., Benedetto, A., and Rattin, J. Increase of spinach growth through the use of larger plug cell volume and an exogenous BAP spray. *Journal of Experimental Agriculture*. 2015, 6(6):372–383. <https://doi.org/10.9734/AJEA/2015/14979>
36. Dong, Y., Zheng, Z., Yan, J., and Deng, S. Natural degradation simulation and phytoremediation of oil-contaminated soil. *In IOP Conference Series: Earth and Environmental Science*. 2021, (680):1-8. [doi/10.1088/1755-1315/680/1/012115](https://doi.org/10.1088/1755-1315/680/1/012115)
37. Eiró, L., Ferreira, M., Frazão, D., Aragão, W., Souza-Rodrigues, R., Fagundes, N., Maia, L., and Lima, R. Lead exposure and its association with neurological damage: systematic review and meta-analysis. *Environmental Science and Pollution Research*. 2021, 28:37001–37015. <https://doi.org/10.1007/s11356-021-13536-y>
38. Elliott, H., and Brown, G. Comparative evaluation of NTA and EDTA for extractive decontamination of Pb-polluted soils. *Water, Air, and Soil Pollution*. 1986 (45):3-4. <https://doi.org/10.1007/BF00283464>
39. Environmental Protection Agency (EPA). Assessing Brownfield sites. June 2020.

40. Environmental Protection Agency (EPA). Build Act - Division N of consolidated appropriations act. 2018; 1-19.
41. European Commission – Joint Research Centre Institute for Health and Consumer Protection. European Chemicals Bureau (ECB): edetic acid (EDTA). European Union Risk Assessment Report. 2004, (49): 1-154.
42. Evangelou, M., Ebel, M., and Schaeffer, A. Chelate assisted phytoextraction of heavy metals from soil. Effect, mechanism, toxicity, and fate of chelating agents. *Chemosphere*. 2007, (68):989-1003. <https://doi.org/10.1016/j.chemosphere.2007.01.062>
43. Fasinu, P., and Orisakwe, O. Heavy metal pollution in sub-Saharan Africa and possible implications in cancer epidemiology. *Asian Pacific Journal of Cancer Prevention*. 2013, (14):3393–3402. <https://doi.org/10.7314/apjcp.2013.14.6.3393>
44. Fathabadi, B., Dehghanifiroozabadi, M., Aaseth, J., Sharifzadeh, G., Nakhaee, S., Rajabpour-Sanati, A., Amirabadizadeh, A., and Mehrpour, O. Comparison of blood lead levels in patients with Alzheimer’s disease and healthy people. *American Journal of Alzheimer’s Disease and Other Dementias*. 2018, (33):541–547. <https://doi.org/10.1177/1533317518794032>
45. Fike, J., Pease, J., Owens, V., Farris, R., Hansen, J., Heaton, E., Hong, C., Mayton, H., Mitchell, R., and Viands, D. Switchgrass nitrogen response and estimated production costs on diverse sites. *GCB Bioenergy*. 2017, (9):1526–1542. <https://doi.org/10.1111/gcbb.12444>
46. Filgueiras, A., Lavilla, I., and Bendicho, C. Chemical sequential extraction for metal partitioning in environmental solid samples. *Journal of Environmental Monitoring*. 2002, 4:823–857. <https://doi.org/10.1039/B207574C>

47. Filippelli, G., Laidlaw, M., Latimer, J., and Raftis, R. Urban lead poisoning and medical geology: an unfinished story. *GSA Today*. 2005, (15):4-11. [http://dx.doi.org/10.1130/1052-5173\(2005\)015%3C4:ULPAMG%3E2.0.CO;2](http://dx.doi.org/10.1130/1052-5173(2005)015%3C4:ULPAMG%3E2.0.CO;2)
48. Filippelli, G., and Laidlaw, M. The elephant in the playground: confronting lead-contaminated soils as an important source of lead burdens to urban populations. *Perspectives in Biology and Medicine*. 2010, 53(1): 31–45. <http://doi.org/10.1353/pbm.0.0136>
49. Freitas, E.V., Nascimento, C.W., Souza, A., and Silva, F.B.. Citric acid-assisted phytoextraction of lead: a field experiment. *Chemosphere*. 2013, 92(2):213-217. <https://doi.org/10.1016/j.chemosphere.2013.01.103>
50. Foy, C., Chaney R., and White, M. The physiology of metal toxicity in plants *Annual Review of Plant Physiology*. 1978, (29): 511-566. <https://doi.org/10.1146/annurev.pp.29.060178.002455>
51. Garbisu, C., and Alkorta, I. Phytoextraction: a cost-effective plant-based technology for the removal of metals from the environment. *Bioresource Technology*. 2001, 77(3):229-236. [https://doi.org/10.1016/S0960-8524\(00\)00108-5](https://doi.org/10.1016/S0960-8524(00)00108-5)
52. Gethami, F., and Sayed, H. In vitro: Influence of various concentrations of plant growth regulators (BAP & NAA) and sucrose on regeneration of *Chenopodium quinoa* Willd. plant. *Asian Journal of Biology*. 2020, 9(4):34-43. <https://doi.org/10.9734/ajob/2020/v9i430095>
53. Gilly, A. Chemically induced phytoextraction of lead (Pb) contaminated soil by switchgrass (*Panicum virgatum L.*). Kennesaw State University, Kennesaw, GA, Fall 2020. Master of Science in Integrative Biology Theses.

54. Gomes, H. Phytoremediation for bioenergy: challenges and opportunities. *Environmental Technology Reviews*. 2012, (1)1: 59–66. <https://doi.org/10.1080/09593330.2012.696715>
55. González-Chávez, M., Carrillo-González, R., Cuellar-Sánchez, A., Delgado-Alvarado, A., Suárez-Espinosa, J., Ríos-Leal, E., Solís-Domínguez, F., and Maldonado-Mendoza, I., Phytoremediation assisted by mycorrhizal fungi of a Mexican defunct lead-acid battery recycling site. *Science of The Total Environment*. 2019, 650(2): 3134-3144. <https://doi.org/10.1016/j.scitotenv.2018.10.031>
56. Greipsson, S. Phytoremediation. *Nature Education Knowledge*. 2011, 3(10), 7.
57. Greipsson, S., Tay, C.F., Whatley, A., and Deocampo, D. Sharp decline in lead contamination in topsoil away from a smelter and lead migration in Ultisol. *World Environment*. 2013, 3(3): 102-107. <https://doi:10.5923/j.env.20130303.05>
58. Greipsson, S., Koether, M., and McElroy, T. Effects of supplementary nutrients (soil-nitrogen or foliar-iron) on switchgrass (*Panicum virgatum L.*) grown in Pb-contaminated soil. *Journal of Plant Nutrition*. 2022. <https://doi.org/10.1080/01904167.2022.2068433>
59. Greipsson, S., McElroy, T., Koether, M. Foliar application of salicylic acid and DA-6 on switchgrass (*Panicum virgatum L.*) grown in Pb-contaminated soil; implications for phytoextraction. *Communications in Soil Science and Plant Analysis*. 53(16): 2045 -2053 2022. <https://doi.org/10.1080/00103624.2022.2070193>
60. Hart, G., Koether, M., McElroy T., and Greipsson, S. Evaluation of chelating agents used in phytoextraction by switchgrass of lead (Pb) contaminated soils. *Plants*. 2022, (11) 1012. <https://doi.org/10.3390/plants11081012>

61. He, Z., Shentu, J., Yang, X., Baligar, V., Zhang, T., Stoffella, P. Heavy metal contamination of soils: sources, indicators, and assessment. *Journal of Environmental Indicators* 2015(9):17–18.
62. Health Canada – Environment and climate change Canada. Screening assessment: EDTA and its salts group. 2018; 1-25.
63. Hernández-Allica, J., Becerril, J., and Garbisu, C. Assessment of the phytoextraction potential of high biomass crop plants. *Environmental Pollution*. 2008, 152(1): 32-40.  
<https://doi.org/10.1016/j.envpol.2007.06.002>
64. Hildebrandt U., Regvar, M., and Bothe, H. Arbuscular mycorrhiza and heavy metal tolerance, *Phytochemistry*. 2007, 68(1), 139-146.  
<https://doi.org/10.1016/j.phytochem.2006.09.023>.
65. Hovsepyan, A., and Greipsson, S. Effect of arbuscular mycorrhizal fungi on phytoextraction by corn (*Zea mays*) of lead- contaminated soil. *International Journal of Phytoremediation*. 2004, 6(4):305-321.
66. Hu, X., Liu, X., Zhang, X., Cao, L., Chen, J., and Yu, H. Increased accumulation of Pb and Cd from contaminated soil with *Scirpus triquetus* by the combined application of NTA and APG. *Chemosphere*. 2017, 188, 397–402.  
<https://doi.org/10.1016/j.chemosphere.2017.08.173>
67. Huang, J., Chen, J., Berti, W., and Cunningham, S. Phytoremediation of lead contaminated soils: role of synthetic chelates in lead phytoextraction. *Environmental Science Technology*. 1997, 31, 800-805. <http://dx.doi.org/10.1021/es9604828>
68. Jan, A., Azam, M., Siddiqui, K., Ali, A., Choi, I., and Haq, Q. Heavy metals and human health: mechanistic insight into toxicity and counter defense system of antioxidants.

*International Journal of Molecular Sciences*. 2015, (16), 12, 29592-29630.

<https://doi.org/10.3390/ijms161226183>

69. Johnson, D., Deocampo, D., El-Mayas, H., and Greipsson, S. Induced phytoextraction of lead through chemical manipulation of switchgrass and corn; role of iron supplement.

*International Journal of Phytoremediation*. 2015, 17(12), 1192–1203.

<https://doi.org/10.1080/15226514.2015.1045134>

70. Joosse, T. Nearly half of all bald eagles have lead poisoning: Pollution could hamper population growth. *Plants and Animals - Science*. 2022. doi:10.1126/science.ada1409

71. Khan, J., Malangisha, G.K., Ali, A., Mahmoud, A., Yang, J., Zhang, M., and Hu, Z. Nitric oxide alleviates lead toxicity by inhibiting lead translocation and regulating root growth in watermelon seedlings. *Horticulture Environment and Biotechnology*. 2021.

<https://doi.org/10.1007/s13580-021-00346-x>

72. Khodadoust, A., Reddy, K., and Maturi, K. Effect of different extraction agents on metal and organic contaminant removal from a field soil. *Journal of Hazardous Materials*. 2005

(117)1, 15–24. <https://doi.org/10.1016/j.jhazmat.2004.05.021>

73. Lal, S., Ratna, S., Said, O., and Kumar, R. Biosurfactant and exopolysaccharide-assisted rhizobacterial technique for the remediation of heavy metal contaminated soil: an advancement in metal phytoremediation technology. *Environmental Technology and Innovation*. 2018, (10): 234-263.

<https://doi.org/10.1016/j.eti.2018.02.011>

74. Lee, J. Kaunda, R., Sinkala, T., Workman, C., Bazilian, M., and Clough, G.

Phytoremediation and phytoextraction in Sub-Saharan Africa: addressing economic and social challenges. *Ecotoxicology and Environmental Safety*. 2021, (226);112864.

75. Lee, M., Park J., and Chung, J. Comparison of the lead and copper adsorption capacities of plant source materials and their biochars. *Journal of Environmental Management*. 2019, (236):118-124. <https://doi.org/10.1016/j.jenvman.2019.01.100>
76. Liu, L., Luo, D., Yao, G., Huang, X., Wei, L., Liu, Y., Wu, Q., Mai, X., Liu, G., and Xiao, T. Comparative activation process of Pb, Cd and Tl Using chelating agents from contaminated red soils. *International Journal of Environmental Research and Public Health*. 2020,17(2), 497. <https://doi.org/10.3390/ijerph17020497>
77. Lou, Y., Luo, H., Hu, T., Li, H., and Fu, J. Toxic effects, uptake, and translocation of Cd and Pb in perennial ryegrass. *Ecotoxicology*. 2013, 22(2), 207-214. <https://doi.org/10.1007/s10646-012-1017-x>
78. Lwin, C., Seo, B., Kim, H., Owens, G., and Kim, K. Application of soil amendments to contaminated soils for heavy metal immobilization and improved soil quality—a critical review. *Soil Science and Plant Nutrition*. 2018, 64:2, 156-167. <https://doi.org/10.1080/00380768.2018.1440938>
79. Martinez-Finley, E., Chakraborty, S., Fretham, S., and Aschner, M. Cellular transport and homeostasis of essential and nonessential metals. *Metallomics*. 2012, 4, 593–605. <https://doi.org/10.1039/c2mt00185c>
80. Mazerolle, M. AICcmodavg: model selection and multimodel inference based on (Q)AIC(c). 2020, R package version 2.3-1. <https://cran.r-project.org/package=AICcmodavg>.
81. McLaughlin, S., and Kszos, L. Development of switchgrass (*Panicum virgatum*) as a bioenergy feedstock in the United States. *Biomass and Bioenergy*. 2005, 28(6), 515-535. <https://doi.org/10.1016/j.biombioe.2004.05.006>

82. McGrath, S., and Zhao, F. Phytoextraction of metals and metalloids from contaminated soils. *Current Opinion in Biotechnology*. 2003, 14(3): 277-282.  
[https://doi.org/10.1016/s0958-1669\(03\)00060-0](https://doi.org/10.1016/s0958-1669(03)00060-0)
83. McFarland, M., Hauer, M., and Reuben, A. Half of US population exposed to adverse lead levels in early childhood. *Proceedings of the National Academy of Sciences*. 2022, (119)11:1-7. <https://doi.org/10.1073/pnas.2118631119>
84. Meers, E., and Tack, F. The potential of foliar treatments for enhanced phytoextraction of heavy metals from contaminated soil. *Remediation Journal*. 2004, 14:111-123.  
<https://doi.org/10.1002/rem.20025>
85. Melo, É., Nascimento, C., Accioly, A., and Santos, A. Phytoextraction and fractionation of heavy metals in soil after multiple applications of natural chelants. *Scientia Agricola*. 2008, 65(1): 61-68. <https://doi.org/10.1590/S0103-90162008000100009>
86. Min, D., Guragain, Y., Prasad, V., Vadlani, P. and Lee, J. Effects of different genotypes of switchgrass as a bioenergy crop on yield components and bioconversion potential. *Journal of Sustainable Bioenergy Systems*. 2017, 7: 27-35. <https://doi.org/10.4236/jsbs.2017.71003>
87. Mufalo, W., Tangviroon, P., Igarashi, T. Ito, M., Sato, T., Chirwa, M., Nyambe, I., Nakata, H., Nakayama, S., and Ishizuka, M. Solid-phase partitioning, and leaching behavior of Pb and Zn from playground soils in Kabwe, Zambia. *Toxics*. 2021, 9(10) 248:1-14.  
<https://doi.org/10.3390/toxics9100248>
88. Namdjoyan, S., and Kermanian, H. Exogenous nitric oxide (as sodium nitroprusside) ameliorates arsenic-induced oxidative stress in watercress (*Nasturtium officinale* R. Br.) plants. *Scientia Horticulturae* 2013, 161: 350–356

89. Nazir, S., Gomathinayagam, S., and Ansari, A. The use of *Spirodela polyrrhiza* (duckweed) and *Eichhornia crassipes* (water hyacinth) to phytoremediate wastewater in Guyana. *Asian Journal of Applied Science and Technology*. 2020, 4(3): 69-75.  
<http://doi.org/10.38177/ajast.2020.4310>
90. Neill, S.J., Desikan, R., Clarke, A., Hurst, R.D., and Hancock, J.T. Hydrogen peroxide and nitric oxide as signaling molecules in plants. *Journal of Experimental Botany*. 2002, 53: 1237–1247
91. Nevin, R. How lead exposure relates to temporal changes in IQ, violent crime, and unwed pregnancy. *Environ. Res.* 2000, 83: 1–22. <https://doi.org/10.1006/enrs.1999.4045>
92. Nguyen, H., Lyttek, E., Lal, P., Wiczerak, T., and Burli, P. Assessment of switchgrass-based bioenergy supply using GIS-based fuzzy logic and network optimization in Missouri (U.S.A.). *Energies*. 2020, 13, 4516. <https://doi.org/10.3390/en13174516>
93. Nissim, W., and Labrecque, M. Reclamation of urban brownfields through phytoremediation: implications for building sustainable and resilient towns. *Urban Forestry and Urban Greening*. 2021, (65):127365
94. Oviedo, C., and Rodríguez, J. EDTA: the chelating agent under environmental scrutiny. *Química Nova*. 2003, 26(6): 901-905. <https://doi.org/10.1590/S0100-40422003000600020>
95. Paulson, J. A., and Brown, M. The CDC blood lead reference value for children: time for a change. *Environmental Health*. 2019,18, 16. <https://doi.org/10.1186/s12940-019-0457-7>
96. Peraza, M., Ayala-Fierro, F., Barber, D., Casarez, E., and Rael, L. Effects of micronutrients on metal toxicity. *Environmental Health Perspectives*. 1998 (106): 203–216.  
<https://doi.org/10.2307/3433921>

97. Perry, R., El-Mayas, H., Krogstadt E., and Greipsson, S. Chemically enhanced phytoextraction of lead-contaminated soils. *International Journal of Phytoremediation*. 2012, 14, 703-713. <https://doi.org/10.1080/15226514.2011.619236>
98. Phielor, R., Voit, A., and Kothe, E. Microbially supported phytoremediation of heavy metal contaminated soils: strategies and applications. *Advances in Biochemical Engineering/ Biotechnology*. 2014,141, 211-235. [https://doi.org/10.1007/10\\_2013\\_200](https://doi.org/10.1007/10_2013_200)
99. Quarshie, S., Xiao, X., and Zhang, L. Enhanced phytoremediation of soil heavy metal pollution and commercial utilization of harvested plant biomass: a review. *Water Air and Soil Pollution*. 2021, 232, 475 . <https://doi.org/10.1007/s11270-021-05430-7>
100. Ranieri, E., D’Onghia, G., Ranieri, F., Petrella, A., Spagnolo, V., and Ranieri, A. Phytoextraction of Cr (VI)-contaminated soil by *Phyllostachys pubescens*: a case study. *Toxics*. 2021 (9): 312, 1-15. <https://doi.org/10.3390/toxics9110312>
101. Robinson, D., Hayes, A., and Couch, S. Broom: convert statistical objects into tidy tibbles. 2021, R package version 0.7.10. <https://CRAN.R-project.org/package=broom>
102. RStudio Team. RStudio: Integrated development environment for R. 2021, RStudio, PBC, Boston, MA <http://www.rstudio.com/>.
103. Sarkar, A., Wang, Q., Asaeda, T., and Kaneko, Y. Arbuscular mycorrhiza confers lead tolerance and uptake in *Miscanthus sacchariflorus*. *Chemistry and Ecology*. 2018, (34), 5, 454–469. <https://doi.org/10.1080/02757540.2018.1437150>
104. Sharma, A., Sidhu, G.P.S., Araniti, F., Bali, A. S., Shazad, B., Tripathi, D. K., Brestic M., Skalicky, M., and Landi, M. The role of salicylic acid in plants exposed to heavy metals. *Molecules*. 2020, 25(3):540, 1-22. <https://doi.org/10.3390/molecules25030540>

105. Shen, Z.G., Li, X.D., Wang, C.C., Chen, H.M., and Chua, H. Lead phytoextraction from contaminated soil with high-biomass plant species. *Journal of Environmental Quality*. 2002, 31(6), 1893-1900. <https://doi.org/10.2134/jeq2002.1893>
106. Shinta, Y.C., Zaman, B., and Sumiyati, S. Citric acid and EDTA as chelating agents in phytoremediation of heavy metals in polluted soil: a review. *IOP Conf. Ser.: Earth Environmental Science*. 2021, 896, 012023. doi:10.1088/1755-1315/896/1/012023
107. Shrestha P., Bellitürk, K., and Görres, J. Phytoremediation of heavy metal-contaminated soil by switchgrass: a comparative study utilizing different composts and coir fiber on pollution remediation, plant productivity, and nutrient leaching. *International Journal of Environmental Research and Public Health*. 2019, 16, 7, 1261. <https://doi.org/10.3390/ijerph16071261>
108. Skerfving, S., Löfmark, L., Lundh, T., Mikoczy, Z., and Strömberg, U. Late effects of low blood lead concentrations in children on school performance and cognitive functions. *Neurotoxicology*. 2015, 49, 114–120. <https://doi.org/10.1016/j.neuro.2015.05.009>
109. Steliga, T., and Kluk, D. Assessment of the suitability of *Melilotus officinalis* for phytoremediation of soil contaminated with petroleum hydrocarbons (TPH and PAH), Zn, Pb and Cd Based on toxicological tests. *Toxics*. 2021, 9,(7): 148, 1-29. <https://doi.org/10.3390/toxics9070148>
110. Tangahu, B., Abdullah, S., Basri, H., Anaur, N., and Mukhlisin, M. A review on heavy metals (As, Pb, and Hg) uptake by plants through phytoremediation. *International Journal of Chemical Engineering*, 2011, 1–31. <https://doi.org/10.1155/2011/939161>
111. U.S. EPA. Method 3050B: Acid digestion of sediments, sludges, and soils, revision 2; U.S. EPA: Washington, DC, USA, 1996.

112. Van der Heijden, M., Martin, F., Selosse, M. and Sanders, I. Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist*. 2015, 205: 1406–1423. <https://doi.org/10.1111/nph.13288>
113. Varshney, S., Hayat, S., Alyemeni, M., and Ahmad, A. Effects of herbicide applications in wheat fields: is phytohormones application a remedy? *Plant Signaling & Behavior*. 2012, 7(5), 570–575. <https://doi.org/10.4161%2Fpsb.19689>
114. Vicente, M., and Plasencia, J. Salicylic acid beyond defense: its role in plant growth and development. *Journal of Experimental Botany*. 2011 (62), 10, 3321–3338. <https://doi.org/10.1093/jxb/err031>
115. Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L., Francois, R., Grolemund, G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T., Miller, E., Bache, S., Mueller, K., Ooms, J., Robinson, D., Seidel, D., Spinu, V., Takahashi, K., Vaughan, D., Wilke, C., Woo, K., and Yutani, H. Welcome to the tidyverse. *Journal of Open Source Software*. 2019, 4(43), 1686 1-6. <https://doi.org/10.21105/joss.01686>
116. Wickham, H. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York, 2016. Alboukadel Kassambara . ggpubr: 'ggplot2' Based Publication Ready Plots. 2020, R package version 0.4.0. <https://CRAN.R-project.org/package=ggpubr>
117. Wu, X., Cobbina, S., Mao, G., Xu, H., Zhang, Z., Yang, L. A review of toxicity and mechanisms of individual and mixtures of heavy metals in the environment. *Environmental Science and Pollution Research*. 2016, 23(9), 8244–8259. <https://doi.org/10.1007/s11356-016-6333-x>
118. Xu, Z., Ban, Y., Li, Z., Hui, C., Yang, R., and Tang, M. Arbuscular mycorrhizal fungi play a role in protecting roots of *Sophora viciifolia* Hance. from Pb damage associated with

- increased phytochelatin synthase gene expression. *Environmental Science and Pollution Research*. 2014, (21), 12671–12683. <https://doi.org/10.1007/s11356-014-3209-9>
119. Yan, A., Wang, Y., Tan, S.N., Mohd Yusof M., Ghosh, S., and Chen, Z. Phytoremediation: a promising approach for revegetation of heavy metal-polluted land. *Frontiers in Plant Science*. 2020, 11:359. <https://doi.org/10.3389/fpls.2020.00359>
120. Yang, Y., Lian, Y., Han, X., Chiu, T., Ghosh, A., Chen, H., and Tang, M. The roles of arbuscular mycorrhizal fungi (AMF) in phytoremediation and tree-herb interactions in Pb contaminated soil. *Scientific Reports*. 2016, 6(1). <https://dx.doi.org/10.1038%2Fsrep20469>
121. Zhuang, P., Zou, B., Li, N., and Li, Z. Heavy metal contamination in soils and food crops around Dabaoshan mine in Guangdong, China: implication for human health. *Environmental Geochemistry and Health*. 2006, 31(6):707-715. <https://doi.org/10.1007/s10653-009-9248-3>
122. Zimdahl, R., and Skogerboe, R. Behavior of lead in soil. *Environmental Science and Technology*. 1977, 11(13), 1202-1207. <https://doi.org/10.1021/es60136a004>
123. Zottini, M., Costa, A., Michele, R., Ruzzene, M., Carimi, F., Schiavo, F. Salicylic acid activates nitric oxide synthesis in *Arabidopsis*. *Journal of Experimental Botany*. 2007, (58)6:1397-14050. <https://doi.org/10.1093/jxb/erm001>